

ALLELOPATHIC POTENTIAL OF ASARUM EUROPAEUM TOWARD LYCOPERSICON ESCULENTUM

Monica MARIAN^{*}, Camelia NICULA, Anca PETER, Georgeta MATEI, Codruta SPATAR North University of Baia Mare, Romania

ABSTRACT. Asarum europaeum L. contains water-soluble substances which manifest allelopathic potential. Aqueous extracts from leaves and stems of *Asarum europaeum* were assayed to determine their allelopathic effects on *Lycopersicon esculentum* and *Zea mays* seeds germination and early seedling growth. The germination of the investigated seeds was found to be inhibited with increasing of the *Asarum europaeum* L. extract concentration. Moreover, the active substances extracted from leaves were found to be inhibitorier on the seeds germination in comparison with those extracted from stems.

KEYWORDS: Allelopathy, Asarum europaeum, leaves, rhizomes, growth inhibitor

INTRODUCTION

Allelopathy (Molisch, 1937) is the chemical interaction between plants, including stimulatory as well as inhibitory influences. Allelopathy plays an important role in both natural and agro-ecosystems and has potential in integrated weed management. Plants contain thousands of natural products, but not all are supposed as having allelopathic effects (Bell et al., 1980; Rice, 1984). Allelochemicals are low molecular weight compounds excreted from plants during the processes of secondary metabolism (Rice, 1992) and they can accumulate in plants, soils and other organisms. These compounds vary in chemical composition, concentration and localization in plant tissues and from plant to plant with changes in both biotic and abiotic conditions (Inderjit et al., 2003). Asarum europaeum, commonly known as Asarabacca, European Wild Ginger, Haselwort, and Wild Spikenard, is a species of wild ginger with single axillary dull purple flowers, lying on the ground. It is widespread across Europe, ranging from southern Finland and northern Russia down to southern France, Italy and the Republic of Macedonia. It is also grown extensively outside its natural habitat as ornamental plant. It is sometimes harvested for use as a spice or as source of flavors (Seidemann, 2005; Katzer, 2010). The species of the Asarum genus contain a high variety of chemical compounds including flavonoids such chalcone, flavonols, anthocyanides, methylisoeugenol, α -asarone (19.2%), α -asarone and methyleugenol. Iwashina et al. have extracted and isolated from the Asarum genus two new chalcone glycosides, 2`,4`-di-Oglucoside chalcononaringenin and chalcononaringenin 2⁻O-glucoside-4⁻Ogentiobioside, from the leaves of A. canadense with seven known flavonol glycosides, quercetin 3-Ogalactoside, quercetin 3-O-robinobioside, quercetin 3-O-galactoside-7-O-rhamnoside, kaempferol 3-0galactoside, kaempferol 3-O-glucoside, kaempferol 3-Ogalactoside- 7-O-rhamnoside and isorhamnetin 3-Ohamnosylgalactoside (Iwashina et al, 2002). Flavonoids are generally considered to inhibit germination and cell growth (Berhow et.al, 1999), thus their allelochemical

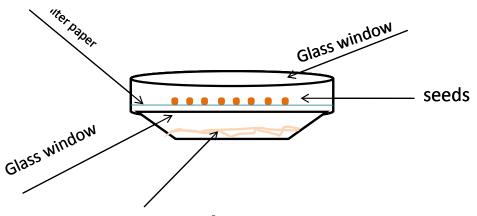
release early in the season could affect other species at susceptible life stages (e.g., germinating seeds and young seedlings) (Weir et al., 2003). In addition to temporal and seasonal variation in allelochemical production, differences may exist among genotypes, populations, or plants of different ages. On the other hand, it was demonstrated that the essential oils from -Asarum have antimicrobial activity (Shunying et al., 2006). In previous investigations, this essential oil was found to possess the promising antifungal activity against a variety of plant pathogens (Liu et al., 2007; Wang and Ji, 2007; Wang et al., 2008, Yang Dan et al., 2010). This article aims to identify the allelopathical effect of Asarum europaeum and moreover to compare the inhibitory effect of the different vegetative organ extract on the germination and seedling growth of Lycopersicon esculentum and Zea mays.

MATERIAL AND METHODS

Obtaining of the extracts from rhizomes and leaves of Asarum europaeum

In March 2010, about a hundred samples of Asarum were randomly taken from Fagetum and Carpinetum forests near Baia Mare and alongside the Somes river. The plant tissues were clipped by hand 1 cm above the soil and directly oven-dried at 60°C for 5 days. Forty grams of dried leaves, respectively rhizomes, were extracted by soaking in 1 l-distilled water at 24^oC for 24 h in a stirrer Heidolpf Unimax 1010 to give a concentration of 40 g dry tissue /L. The extract was filtered using an Laboport vacuum pump. Each stock extract was diluted appropriately with sterile distilled water 1:1, 1:2 and 1:3 (v:v). Distilled water was used as reference. The seeds of the species tested, namely tomato (Lycopersicon esculentum L.) and maize (Zea mays L.), were used for germination assays. The seeds were sterilized with 5% sodium hypochlorite for 10 min and five times rinsed with distilled water. In each experiment 100 seedlings were used and the experiments were repeated three times. To evaluate the allelopathic effect to water soluble extracts, 50 seeds of tomato and separately maize, placed in a plate, containing two layers of filter paper moistened with 10

ml of aqueous extract of rhizomes and leaves, respectively, in different dilutions, as Figure 1 suggests. The plates were maintained, for 7 days, in a Sanyo environmental test chamber (temperature 23^{0} C, illumination 10.000 lx and humidity 82%). The experiments were replicated three times.



Aqueous extract of Asarum europaeum

Fig.1. Scheme of the germination experiments

Table 1.

Parameters of germination and seedling growth of tomato seeds in the presence of rhizomes and leaves extracts of *Asarum europaeum*

	Time (days)											
	3	4	5	6	7	15	32	39				
	Germination (number of germinated seeds) (%)						Plant growth (cm)					
Rhizomes ex	tract dilution	(v:v)										
1 :1	4	11	31	42	57	0,7	3	6				
1 :2	6	15	38	45	52	0,8	4	7				
1 :3	12	18	36	41	55	1,3	5,5	8				
Leaves extra	ct dilution (v:	v)										
1 :1	0	2	13	25	41	0,2	3	5				
1 :2	7	16	35	41	56	0,3	5	6				
water	50	64	67	69	75	1,5	7	11				

Table 2.

Parameters of germination and seedling growth of maize seeds in the presence of rhizomes and leaves extracts of *Asarum europaeum*

	Time (days)								
	3	8	15	20	32	39			
	Germination (n	umber of germinat		Plant growth (cm)					
Rhizomes extract dilu		C C			Ū	. ,			
1 :1	60	70	100	0,8	3,2	5			
1 :2	80	86	100	0,9	5	6,2			
1 :3	82	94	100	1,5	7,3	8,4			
Leaves extract dilutio	n (v:v)								
undilluted	60	76	83						
1 :1	36	38	60	1:1	4	11			
1 :2	72	80	82	1:2	6	15			
water	92	100	100	1:3	12	18			

RESULTS AND DISCUSSION

Asarum europaeum is a species peculiar to the *Querco-Fagetea* genus, populating most of the forests of this genus, adding up into facieses and appearing as a competitor that visibly contributes to the extinction of the other species. One of the reasons is the low height of this species, its repent and reniform shapes covering high areas. The allelopathic influence that such species exercise on the germination process of other plants, as well as on those plants growing process could be

another explanation. This species is known for the ability to preserve green leaves during winter and the rejuvenation of those leaves occurs only in spring, relatively at the same time with the germination of the other species on the same soil. Because of these reasons we have tested the allelopathic capacity of rhizomes and separately the capacity of the *asarum europaeum* leaves on a reference sample of plants. We have considered for our experiments both a dicotyledonuos and a monocotyledonous species. In order to test the germination capacity we have performed separate experiments. We found a germination capacity in the 80% - 100% range for both species

Details in table 1 and in fig. 2, 6 and 8 reveal for the rhizome extract a different allelopathic capacity as compared to that of leaves on the germination of la *Lycopersicon esculentum*. Both the rhizome extract and the extract of leaves, at various dilutions, inhibit the germination capacity with almost 50% as compared to the witness sample tested in distilled water. This proves the unquestionable existence of an adaptive mechanism that the plant uses against other species. The allelopathic effect of the species tested shows up particularly as a mechanism that delays the germination of other species, in a way that debilitates the species in the neighbourhood of *Asarum europaeum*.

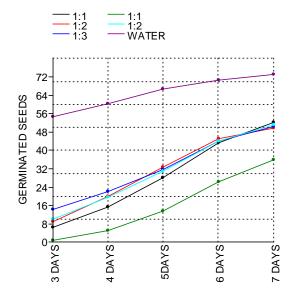


Fig. 2. No. of germinated seeds of Lycopersicon esculentum

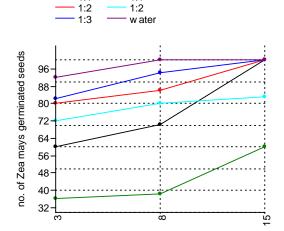


Fig. 4. No. of germinated seeds of Zea mays

The extract of leaves (Fig. 2, the green line) has an inhibitory effect on germination which is obviously stronger than the rhizome extract. The inhibiting effect declines for both the leaves extract and the rhizome extract as the degree of dilution grows, and even for a 1:3 dilution ratio the number of seeds germinated is considerably below the level of the witness sample. Both categories of extracts inhibit the germination of *Zea mays*, (Fig. 4) but while the rhizome only generates a delay, the *Asarum* leaves display an "aggressive" allelopathic effect which strongly inhibits the germination of maize berries. The impact on seedlings is similar with the impact on germination. Leaves have a much stronger inhibitory impact than the rhizome.

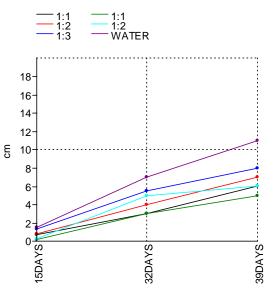


Fig. 3. Lycopersicon esculentum growth

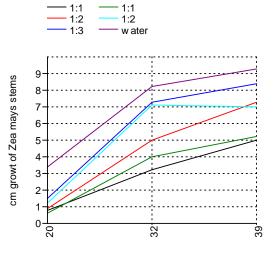


Fig.5. Zea mays growth

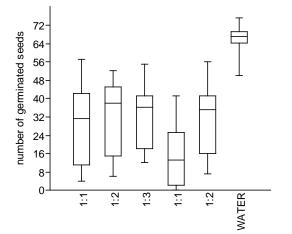


Fig. 6. Average germination in different experimental variants

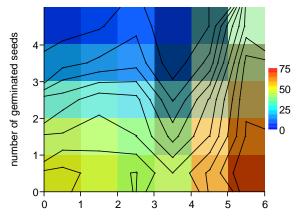


Fig. 8 Germinated Seeds Of Lycopersicon esculentum

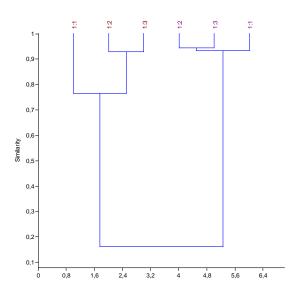


Fig. 10 Cluster analysis between seeds germination of *Lycopersicon & Zea* at different dilution of rhizomes extract (according Euclidian distance)

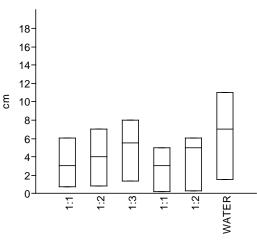


Fig. 7. Mean of seedlings growth in different experimental variants

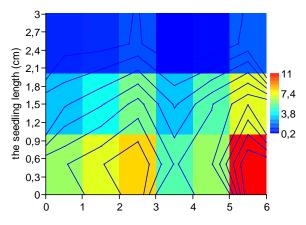


Fig. 9 The stem length of *Lycopersicon esculentum* seedling

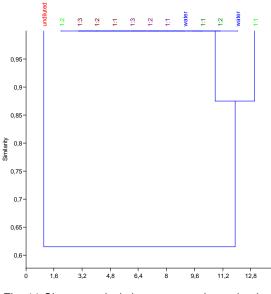


Fig. 11 Cluster analysis between seed germination and plantlet growth at different dilution of *Asarum* extracts (according Jaccard index) Light green – *Lycopersicon plantlets; dark green* – *Zea mays plantlets; purple* – *Lycopersicon seeds; brown* – *Zea mays seeds*

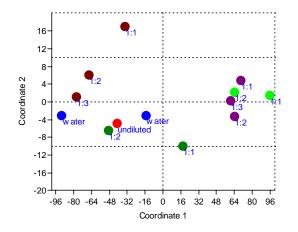


Fig.12. PCO Scatter diagram

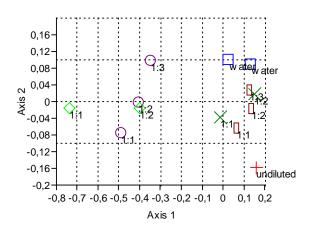


Fig. 13. Correspondence analysis diagram

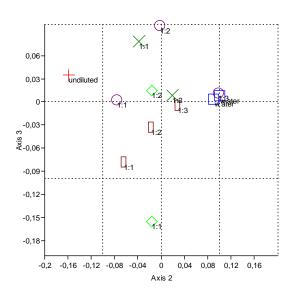


Fig. 14 The analysis of a number of key factors

We have statistically processed the results of the allelopathic effect on the germination material, on Lycopersicon and respectively on Zea mays seedlings (Fig. 10-13). Significant differences came out. The Euclidean similarity index comparing the response of the two species at different dilutions of the rhizome extract is less than one. This shows the existence of different mechanisms against the inhibitory effect (Fig. 10). Maize berries are less sensitive to the allelopathic effect, and even if with a delay, their germination is almost complete, in contrast with the Lycopersicon esculentum seeds on which the inhibitory effect is visible in a much higher proportion. Lycopersicon has against Asarum europaeum's allelopathic activity a different level of resistibility as compared to Zea may. This comes out in (Fig. 11), showing the cluster

analysis for germination, respectively the growth at various degrees of dilution. On one hand, there is very little similarity between the witness samples tested at different degrees of dilution and the undiluted extract of sprouts. As the graph in figure 12 shows, the points that are the closest are those corresponding to samples that belong to the same species and not those corresponding to samples tested at the same dilution. The PCO, the correspondence analysis and the Detrending correspondence analysis (Fig. 12, 13), reveal all the considerable differences among the samples tested. Each of these tests suggests a high degree of dispersion in terms of the response of the samples considered. The samples make up groups depending on the species and on the degree of dilution. The analysis of a number of key factors reveals (Fig.14) that the gap is small between the sample tested with *Zea Mays* berries, undiluted *Asarum* seedlings extract and the witness samples for germination. This proves that immature seedlings hold in little quantity compounds with allelopathic properties. On the other hand, the concentration of active compounds grows as the leaves grow up. This supports the hypothesis that leaves concentrate the highest proportion of active compounds in charge with allelopathic activities.

To conclude, the *Asarum europaeum* species rely on bio-chemical mechanisms to survive and to compete by means of biologically – active compounds with allelopathic effect. These compunds accumulate prevalently in leaves and to a smaller extent in rhizomes. The allelopathic effect takes the form of a delaying mechanism, for monocotyledonous palnts and the form of innhibitory effect for dicotyledonous plants. Inhibition occurs in terms of germination, and in terms of the growth process as well. The

species that are "receptive" to allelopathic compounds develop various resitance mechansims, conferring to plants different sensitivirties. It is demonstrated by relevant experiments that monocotyledonuous plants are more resilient against the inhibitory effect on germination and on the growth process. We think that gaining more insight on the phenomena of biochemical inhibition and also on the ways to counteract inhibitory activities will allow for a better understanding of ecology – specific competition and survival mechanisms within phytoconoses. Revealing details about the biology of plants with potential to be applied in agriculture and forests management is a key outcome of this research exercise.

REFERENCES

- Alforda, E., R., Perryb, Laura G., Qinc, B., Vivancoa, J., M., Paschke, M., W. 2007, A putative allelopathic agent of Russian knapweed occurs in invaded soils, Soil Biology & Biochemistry 39, 1812–1815
- Bell, E.A., Charlwood, B.V., 1980. Secondary plant products. In: Encyclopedia of Plant Physiology, New Series, Springer–Verlag, New York, vol. 8, p. 674.
- Berhow, M.A., Vaughn, S.F., 1999. Higher plant flavonoids: biosynthesis and chemical ecology.
 In: Inderjit, Dakshini, K.M.M., Foy, C.L. (Eds.), Principles and Practices in Plant Ecology: Allelochemical Interactions. CRC Press LLC, Boca Raton, FL, pp. 423–438.
- Inderjit, Callaway, R.M., 2003. Experimental designs for the study of allelopathy. Plant Soil 256, 1– 11.
- Liu, H.Y., Gao, W.W., Fan, Y., Chen, S.L., 2007. Inhibitory effect of essential oil from Asarum heterotropoides Fr. Schmidt var. mandshuricum (Maxim.) Kitag against plant pathogenic fungi. Acta Phytopathol. Sin. 37, 95–98.
- Molisch, H., 1937. Der Einfluss einer Pflanze auf die andere-Allelopathie. Fischer, Jena, Germany.

- Rice, E.L., 1992. Allelopathic effects on nitrogene cycling. In: Rivzi, S.J.H., Rizvi, V. (Eds.), Allelopathy: Basic and Applied Aspects. Chapman & Hall, London, pp. 31–58. (July 2005), World Spice Plants: Economic Usage, Botany, Taxonomy (1 ed.), Germany: Springer, p. 57, ISBN 3540222790.
- Seidemann, Johannes (July 2005), World Spice Plants: Economic Usage, Botany, Taxonomy (1 ed.), Germany: Springer, p. 57, ISBN 3540222790
- ZHU Shunying, YANG Yang, YU Huaidong, YING Yue, LONG Dong, ZOU Guolin, Chemical Composition and Antimicrobial Activity of Essential Oil of Asarum caulescene, 2006, WUJHS Wuhan University Journal of Natural Sciences, Vol. 11 No. 3, 699-703.
- Weir, T.L., Bais, H.P., Vivanco, J.M., 2003. Intraspecific and interspecific interactions mediated by a phytotoxin, (-)-catechin, secreted by the roots of Centaurea maculosa (spotted knapweed). Journal of Chemical Ecology 29, 2397–2412.
- Wang, G.Q., Ji, L.Z., 2007. The antibacterial effects of asarum essential oils on diseases in Ligneous flower in vitro. Northern Hortic. 2007 (9), 220– 222.
- Wang, G.Q., Zhang, J.H., Zhang, M., Ji, L.Z., 2008. The antibacterial effects of Asarum heterotropoides extracts to Pestaloliopsis sp. in vitro. Henan Agr. Sci. 3, 60–63.
- Yang Dan, Hai-Yan Liu, Wei-Wei Gao*, Shi-Lin Chen, 2010, Activities of essential oils from Asarum heterotropoides var. mandshuricum against five phytopathogens, Crop Protection 29 (2010) 295–299